

# Animal Safety Test of *Bacillus thuringiensis* BT Protein

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**Abstract** [Objectives] To determine the biological safety of BT protein from *Bacillus thuringiensis* (Bt) fermentation broth to mammals at high doses. [Methods] Healthy mice were randomly divided into 4 groups with 10 mice in each group. The experimental groups were fed with Bt fermentation supernatant at 0.2, 0.6 and 1.0 mL/kg, respectively, once a day for 7 consecutive days. The blank control group was fed normally without intragastric administration. [Results] There was no significant difference in blood routine and blood biochemical analysis between the experimental group and the control group. After intragastric administration, the mice were dissected, and no obvious pathological changes were found; the heart, liver, spleen, lung and kidney were taken to make tissue sections, and no pathological changes were found by microscopic observation. [Conclusions] Mice can tolerate high doses of BT protein from *B. thuringiensis* fermentation broth.

**Key words** *Bacillus thuringiensis* (Bt), BT protein, Biological safety

## 1 Introduction

*Bacillus thuringiensis* (Bt) can produce exocrine VIP protein in the vegetative growth stage, and a special parasporal crystal CRY protein will be produced in the spore after entering the budding reproductive stage. These proteins, collectively known as BT protein, have insecticidal effects<sup>[1]</sup>. The insecticidal mechanism of BT protein has been widely studied. The basic pathway is that there is a specific site in the midgut epithelial cells of the parasite, which can cause membrane perforation after binding with BT protein, eventually leading to the death of the parasite<sup>[2–4]</sup>. This mechanism mainly acts on lepidopteran insects. Because there is no BT protein receptor on the surface of intestinal cells of mammals, humans and livestock are not sensitive to this protein. Therefore, BT protein is widely used as a harmless insecticide in the field of insecticide<sup>[5–6]</sup>. However, whether the toxicological effects of BT protein will cause pathological changes in mammals due to its accumulation in the body is still uncertain. In view of these, we intended to increase the concentration of *B. thuringiensis*, selected healthy mice as target animals, to understand the effects of BT protein purification liquid on clinical manifestations, physiological and biochemical indicators of animals, so as to provide a theoretical basis for the clinical application of the product.

## 2 Materials and methods

**2.1 Materials and instruments** Forty female healthy mice (experimental animal license: SYXK (Beijing) 2020-0053), weight 18–22 g; BT protein purified solution of *B. thuringiensis*

fermentation, Dingzheng Xinxing Biotechnology (Tianjin) Co., Ltd.; peptone and yeast powder, Tianjin Beilian Fine Chemicals Development Co., Ltd.

SHY-2A Thermostatic Water Bath Shaker, Shanghai Bilon Instrument Manufacturing Co., Ltd.; H1850R Refrigerated Centrifuge, Hunan Xiangyi Laboratory Instrument Development Co., Ltd.; Countess 3, Thermo Fisher Scientific; FDC NX500iVC Blood Biochemical Analyzer, Fuji Film Medical Systems Co., Ltd.; HS-3315 tissue microtome, Beijing Leibo Ruijie Technology Co., Ltd.; ECLIPSE SI microscope, Nikon Precision Machinery (Shanghai) Co., Ltd.

### 2.2 Methods

**2.2.1 Grouping of experimental animals.** The experimental mice were randomly divided into 4 groups with 10 mice in each group, and the grouping is shown in Table 1.

**Table 1** Grouping and treatment of experimental animals

Group	Animals//pcs	Experiment treatment
CK	10	Normal feeding, no drug administration
A1	10	The drug was administered once a day for 7 consecutive days at a dose of 0.2 mL/kg.
A2	10	The drug was administered once a day for 7 consecutive days at a dose of 0.6 mL/kg.
A3	10	The drug was administered once a day for 7 consecutive days at a dose of 1.0 mL/kg.

**2.2.2 Observation indicators.** Generally, the observation period of animal safety experiment is 7–28 d. In this experiment, it is considered that the storage period of fermented products should not be too long, and the longer the storage period, the higher the risk of contamination. In addition, it is considered that BT protein is easily absorbed in the gastrointestinal environment, and 7 d is enough to produce enough accumulation, so it is determined to ob-

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serve the relevant indicators on the 7<sup>th</sup> day.

(i) Clinical observation. During the test, the animals were observed daily for drug-related adverse reactions, such as respiratory and behavioral abnormalities, mental depression, and abnormal defecation.

(ii) Body weight. Weighed the mice before and after the experiment and made records.

(iii) Blood sample analysis. First, blood collection and treatment methods. Blood samples were collected from all mice before and after administration, and placed in two tubes; one tube was a centrifuge tube with anticoagulant for blood routine analysis, and the other tube was a centrifuge tube without anticoagulant. Blood samples were kept at room temperature for 2 h, centrifuged at 2 500 r/min for 5 min, and the upper serum was taken and stored at -20 °C for blood biochemical analysis.

Second, blood routine analysis. White blood cell count (WBC), white blood cell classification[ neutrophil (NE), lymphocyte (LY), monocyte (MO), eosinophil (EO), basophil (BA) ], red blood cell count (RBC), hemoglobin (HGB), hematocrit (HCT), platelet count (PLT) and other indicators were analyzed.

Third, analysis of blood biochemical indicators. Serum alanine aminotransferase (ALT), aspartate aminotransferase (AST), blood urea nitrogen (BUN), creatinine (Cr), albumin (ALB), total protein (TP), total cholesterol (CHO), and triglyceride (TG) were analyzed.

(iv) Anatomical examination. To understand the safety of the preparation in clinical application, the target animals were examined by gross anatomy, and the heart, liver, spleen, lung, kidney and other organs were selected for pathological anatomy and pathological tissue observation.

**2.3 Statistical analysis** The data were presented in mean ±

standard deviation, and the differences between groups were analyzed by chi-square test in SPSS 13.0, with  $P < 0.05$  and  $P < 0.01$  separately indicating significant and extremely significant differences.

3 Results and analysis

**3.1 Clinical observation after administration** The respiration, behavior, spirit, defecation and appetite of the animals in each group were normal after administration, and no adverse reactions occurred.

**3.2 Body weight change before and after administration** The results of animal weight before and after administration are shown in Table 2. The results showed that there was no significant difference in body weight before and after administration ( $P > 0.05$ ).

Table 2 Body weight change of mice before and after administration g

Group	Body weight		Average daily gain
	Before administration	After administration	
A1	19.88 ± 0.79	22.25 ± 2.20	0.34
A2	21.00 ± 0.90	22.48 ± 2.23	0.35
A3	19.83 ± 0.84	22.06 ± 1.66	0.32
CK	19.88 ± 0.73	22.15 ± 1.76	0.32

**3.3 Results of blood routine analysis** The results of blood routine indicators of mice in each group are shown in Table 3. It can be seen from Table 3 that the blood routine indicators before and after administration in each dose group were not significantly different from those in the normal group ( $P > 0.05$ ), indicating that the purified supernatant of Bt fermentation in the dose range of 0.2, 0.6 and 1.0 mL/kg had no significant effect on the blood routine indicators.

Table 3 Effects of different doses of purified supernatant of Bt fermentation on blood routine indicators of mice

Group	WBC// × 10 <sup>9</sup> /L	Classification of white blood cells//%			RBC// × 10 <sup>12</sup> /L	HGB// g/L	HCT//%	PLT// × 10 <sup>9</sup> /L
		NE	LY	MO				
CK	14.72 ± 1.12	38.46 ± 5.17	59.35 ± 5.35	2.00 ± 0.56	6.78 ± 1.00	117.4 ± 14.30	36.17 ± 3.55	426.60 ± 28.05
A1	14.35 ± 1.39	39.53 ± 5.41	58.37 ± 5.48	2.10 ± 0.64	6.70 ± 1.01	109.40 ± 10.74	35.90 ± 3.64	409.80 ± 29.93
A2	15.09 ± 1.48	38.31 ± 6.22	59.85 ± 5.91	1.84 ± 0.68	6.88 ± 1.18	111.64 ± 16.35	35.31 ± 5.12	420.09 ± 32.86
A3	14.41 ± 1.39	40.30 ± 4.43	57.63 ± 4.44	2.07 ± 0.89	6.93 ± 1.01	115.60 ± 14.68	37.18 ± 5.17	423.90 ± 28.95

**3.4 Results of blood biochemical analysis** The detection results of blood biochemical indicators of mice are shown in Table 4. It can be seen from Table 4 that there was no significant difference ( $P > 0.05$ ) between the blood biochemical indicators of each dose

group before and after administration and those of the normal group, indicating that the Bt fermentation and purified supernatant had no significant effect on the blood biochemical indicators in the dose range of 0.2, 0.6 and 1.0 mL/kg.

Table 4 Effects of different doses of purified supernatant of Bt fermentation on blood biochemical indicators of mice

Group	AST//IU/L	ALT//IU/L	BUN//mmol/L	Cr//μmol/L	ALB//g/L	TP//g/L	CHO//mmol/L	TG//mmol/L
CK	110.10 ± 21.93	79.88 ± 10.04	2.17 ± 0.41	168.18 ± 19.34	36.60 ± 6.62	54.60 ± 6.93	1.88 ± 0.13	0.27 ± 0.07
A1	105.70 ± 21.06	76.28 ± 11.59	2.26 ± 0.40	167.16 ± 16.09	36.10 ± 6.33	57.40 ± 5.85	1.80 ± 0.12	0.28 ± 0.07
A2	76.70 ± 18.68	76.60 ± 14.01	2.14 ± 0.40	166.27 ± 16.49	34.40 ± 7.41	55.30 ± 5.74	1.84 ± 0.13	0.25 ± 0.06
A3	107.90 ± 13.89	76.26 ± 13.89	2.30 ± 0.47	165.32 ± 14.50	35.00 ± 5.24	55.10 ± 4.38	1.86 ± 0.20	0.26 ± 0.07

**3.5 Autopsy and histological observation results** The tissue sections of heart, liver, spleen, lung and kidney in the 1.0 mL/kg dose group and the control group are shown in Fig. 1. There were

no obvious macroscopic pathological changes in the results of gross autopsy and histological examination.

# References

- [1] YANG Y. Discussion on efficient cultivation and related management techniques of peach[J]. Contemporary Horticulture, 2017(4): 23. (in Chinese).
- [2] XIONG R, CHEN JY, WANG Y. Study on the flowering-time regulation of outdoor ornamental peaches [J]. Northern Horticulture, 2012(8): 75–77. (in Chinese).
- [3] ZHAO CP, WANG QX, HAN YM, *et al.* Effects of tree shape on the quality of leaf and fruit and the yield in peach[J]. Journal of Northwest A&F University (Natural Science Edition), 2010, 38(6): 160–164, 170. (in Chinese).
- [4] XIAO L, CHEN HJ, DI B, *et al.* The changes in canopy parameters and the effect of yield and quality by two kinds of peach tree shape[J]. Northern Horticulture, 2012(17): 20–23. (in Chinese).
- [5] ZHU GR, WANG LR, CHEN CW, *et al.* A new flat peach cultivar

- ‘Zhongpan 17’ [J]. Journal of Fruit Science, 2020, 37(3): 445–448. (in Chinese).
- [6] WANG ZY, LIU GJ, CHANG RF, *et al.* Breeding of a new late-ripening peach cultivar ‘Qiulian’ [J]. Northern Horticulture, 2020(3): 178–180. (in Chinese).
- [7] YE ZHENG W, SU MS, DU JH, *et al.* A new late ripening yellow peach cultivar ‘Jinshuo’ [J]. Journal of Fruit Science, 2020, 37(3): 441–444. (in Chinese).
- [8] WANG YH, YANG JG, ZHANG JL, *et al.* Experiment and demonstration of green control technology of peach diseases and insect pests in Pinggu, Beijing [J]. China Plant Protection, 2012(8): 20–23. (in Chinese).
- [9] NIU YH, ZHANG T, ZHANG CY, *et al.* Discussion on green control integrated techniques against peach pests [J]. Shaanxi Journal of Agricultural Sciences, 2019, 65(8): 90–92. (in Chinese).

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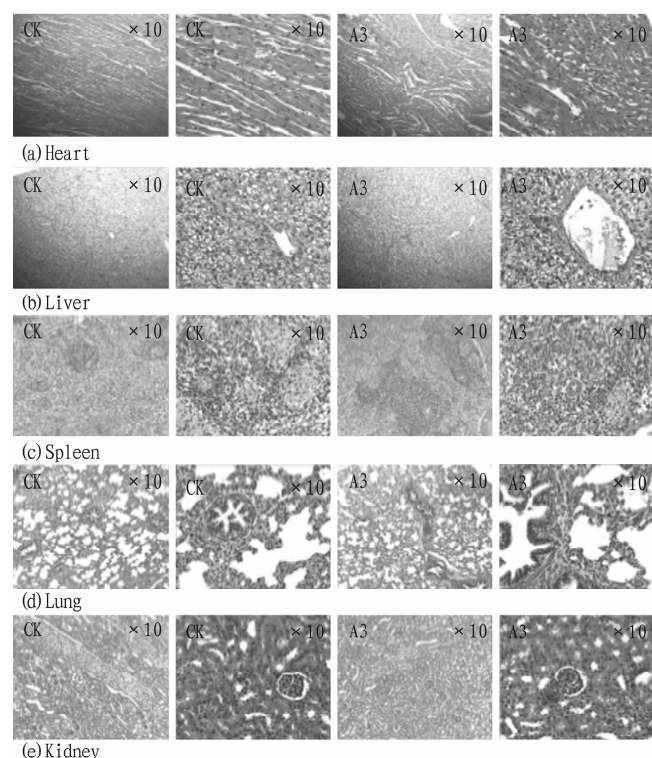


Fig.1 Tissue section

# 4 Conclusions

In summary, the body characteristics of mice were not abnor-

mal compared with the control group when the purified supernatant of Bt fermentation was administered at the doses of 0.2, 0.6 and 1.0 mL/kg. The results of blood routine and blood biochemical analysis showed that there was no significant difference between the two groups. After the administration, mice were dissected and tissue sections were made, and no pathological changes were observed through 10 times and 40 times microscope. The results showed that the purified supernatant of Bt fermentation had high biological safety.

# References

- [1] HUANG HM, ZENG YT, WANG L. Research progress of *Bacillus thuringiensis* and its parasporal crystal insecticide[J]. Guangdong Chemical Industry, 2021, 48(10): 86–87. (in Chinese).
- [2] CHENG GY. Detection and application of water-soluble active substance in fermentation broth of *Bacillus thuringiensis* [D]. Guilin: Guilin University of Technology, 2019. (in Chinese).
- [3] ZHAO KJ, XING YN, HAN LL, *et al.* Optimization of fermentation conditions for *Bacillus thuringiensis* strain Bt20 [J]. Journal of Northeast Agricultural University, 2014, 45(7): 45–53. (in Chinese).
- [4] WANG HR, LIU YW. Research progress in fermentation of Bt biopesticide [J]. Journal of Anhui Agricultural Sciences, 2010, 38(19): 10116–10117. (in Chinese).
- [5] CHEN ZE, WU JX, ZHANG ZG, *et al.* Ten years’ research progress on submerged fermentation of *Bacillus thuringiensis* in China (1990–2000) [J]. Chinese Journal of Biological Control, 2002(1): 33–35. (in Chinese).
- [6] HU F, XU TT, SU XIAN Y, *et al.* Control efficacy of *Bacillus thuringiensis* tiny microgranules on maize lepidopteran pests [J]. Chinese Journal of Biological Control, 2023, 39(1): 46–53. (in Chinese).